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THE SIDE EFFECTS OF ALLOXYDIM SODIUM, SETHOXYDIM, ACIFLUORFEN AND FLUAZIFOP-BUTYL ON LEGUME GROWTH AND NODULATION

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NOTE

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THE SIDE-EFFECTS OF ALLOXYDIM SODIUM, SETHOXYDIM, ACIFLUORFEN AND
FLUAZIFOP-BUTYL ON LEGUME GROWTH AND NODULATION.

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Summary

The effects of four herbicides (alloxydim sodium, sethoxydim, acifluorfen and fluazifop-butyl), were examined on growth and nodulation of five representative leguminous plants; peas, dwarf french beans, field bean, clover and fenugreek.

Effects on plant growth were variable, with factors such as: time of spraying, fertilizer application and sample date all playing a part in this variation.

Alloxydim sodium slightly inhibited pea growth and nodulation, but this effect was minimized when the plants were sprayed at the 6 leaf stage, rather than the 3 leaf stage. Sethoxydim, sprayed at high doses, decreased nodulation in most legume species. Acifluorfen caused a reduction in growth of dwarf french beans and field beans. Nodulation of fenugreek and clover can be severely reduced by fluazifop-butyl, though clover may recover after time.

Introduction

The selectivities of new herbicides are investigated at WRO on a large number of crop and weed species grown in pots. One objective of these studies is to discover crop susceptibilities and to obtain experience of the type of effects produced by each compound.

Amongst the crop species examined are a number of legumes and, since, any effect of herbicides on nodulation could have serious consequences, nodulation is examined visually on legumes treated in all activity and selectivity tests. Any herbicide effecting nodulation in these tests may then be examined in more detail in subsequent experiments. This report describes the results of such experiments on four herbicides.

There are limitations to these experiments. One is that, only one crop variety and one soil type is used. Also plant responses in pot experiments can be very different to those in the field. Thus, the results described should be regarded with caution.

Material and Methods

Soil

A sandy loam soil (Begbroke North), taken from a field at WRO was used for all experiments. The organic matter content is 4.1%, clay content 15% and the pH was 7.0. The soil was sieved (5 mm), mixed with the fertilizers and pesticides shown in Table 1, and placed in 12.5 cm pots.

Fertilizer

Osmocote, controlled release fertilizer, (Sierra Chemicals Europe, Nuverheidsweg 5, Heerlen, Nederland):-

Brown: NPK 15:12:15; short-term release (3-4 months).

Blue: NPK 18:11:10; long-term release (8-9 months).

Vitax QS3, controlled release fertilizer, (Steetly Chemicals Ltd., Liverpool Road North, Burscough, Ormskirk, Lancs L40 0SB) NPK 12:6:5; medium term release (6 months).

Inoculum

In some experiments the plants were inoculated with Rhizobium spp. as shown in Table 2.

A starter culture (100 mls) of Rhizobium spp. was grown in a yeast mannitol liquid medium (Vincent J M, 1970) at 18°C for 5 days with stirring and aeration. This was used to inoculate a 2 litre batch of the same medium and grown for a further 5 days at 18°C, and then mixed with dried, sieved peat to give a moisture content of approximately 50%. The peat inoculum was incubated for a further 5 days at 18°C, and approximately 1 g was added with each seed as it was planted.

Greenhouse Conditions

All experiments were done under greenhouse conditions. During the period October - March additional lighting was provided with sodium lights giving a 14 hour day length. Plants were watered from above as required and when necessary supported with stakes. Maximum, minimum and mean temperatures for each experiment are shown in Table 1.

Spraying

All the herbicides were applied with a laboratory pot sprayer operating at a pressure of 207 kPa (30 lb in²) at a constant speed 30 cm above the plants.

Sampling Procedure

Replicate pots were removed from the greenhouse at the appropriate sampling time. The shoots were cut off at soil level, fresh weights of shoots and pods (if present), were measured. After drying overnight at 105°C the dry weights were also recorded. Roots were removed from the pots and washed to remove the adhering soil, care being taken not to damage or detach any nodules. Greaves M.P., Lockhart L.A. and Richardson W.G. (1978) assessed nodulation in two ways; by excising the nodules and weighing them, and nitrogenase analysis. Neither method proved satisfactory, as the removal of the nodules was excessively time consuming, and the nitrogenase activity varied according to light intensity. In all our experiments the degree of nodulation was assessed visually. The nodulation of the control plants, which were healthy in appearance, unless otherwise stated, was scored as 10 and all treatments scored by comparison. This technique does not permit viable statistical analysis. Root systems were blotted to remove excess water, and fresh and dry weights were recorded. Other experimental details are recorded in Table 2.

Plants grown during tests of herbicide efficacy and selectivity were examined occasionally. Replication in these observations does not permit detailed statistical analysis and thus comments on effects must be viewed with caution.

TABLE 1

FERTILIZER TREATMENTS AND GREENHOUSE TEMPERATURES IN NODULATION EXPERIMENTS

EXPT. NO.	OSMOCOTE	OSMOCOTE	VITAX QS3	MAGNESIUM	5% DDT	SUPERPHOSPHATE	TEMPERATURES (°C)		
	BROWN 15:12:15 g kg ⁻¹ SOIL	BLUE 18:11:10 g kg ⁻¹ SOIL	12:6:5 g kg ⁻¹ SOIL	SULPHATE g kg ⁻¹ SOIL	g kg ⁻¹ SOIL	OF LIME g kg ⁻¹ SOIL	MEAN	MAX.	MIN.
1	1.8	-	-	1.0	0.5	1.2	14	18	8
2	0.2,0.8,3.5	-	-	1.0	0.5	1.2	14	18	8
3	-	-	-	-	-	-	19	29	12
4	-	-	-	1.0	0.5	1.2	19	29	13
5									
Peas	-	-	-	1.0	0.5	1.2			
Dwarf French Bean	-	-	-	1.0	0.5	1.2	19	29	13
Field Bean	-	-	3.0	1.0	0.5	1.2			
6	-	1.8	-	1.0	0.5	1.2	15	23	8
7	-	-	3.0	1.0	0.5	1.2	13	17	8

TABLE 2

PLANT DATA FOR NODULATION EXPERIMENTS

EXP. NO. DATES	SPECIES	CULTIVAR	NO. PLANTS/POT INITIALLY & FINALLY	GROWTH STAGE AT SPRAY	ASSESSMENT WEEKS AFTER SPRAY	INOCULUM	HERBICIDE & DOSE (kg ha ⁻¹)
1 2.10.78- 9.1.79	Pea	Dark Skinned Perfection	5 - 1	3 Leaf & 6 Leaf	3,6,10	<u>Rhizobium leguminosarum</u> RCR 1045	Alloxydim sodium 2
2 12.10.78-10.1.79	Pea	Dark Skinned Perfection	5 - 1	3 Leaf	3,6,10	<u>Rhizobium leguminosarum</u> RCR 1045	Alloxydim sodium 2
3 22.5.79-27.7.79	Pea	Dark Skinned Perfection	5 - 1	3 Leaf 6 Leaf	3,6,8	-	Sethoxydim 0.1,0.4,1.6
4 4.7.79- 3.9.79	Pea	Dark Skinned Perfection	5 - 1	3 Leaf & 6 Leaf	3,5	-	Sethoxydim 1.0
5 4.7.79-18.9.79	Pea	Dark Skinned Perfection	5 - 1	3 Leaf	3,7	-	Sethoxydim 0.4,1.0,1.6
14.9.79-15.11.79	Dwarf french bean	The Prince	3 - 1	2 Unifoliate leaves	3,6	<u>Rhizobium phaseoli</u> RCR 3605	0.4,1.0,1.6
29.11.79- 6.3.80	Field bean	Maris Bead	3 - 1	3 Leaf	4,8	-	0.4,1.0,1.6
6 20.2.79- 1.6.79	Pea	Dark Skinned Perfection	5 - 1	pre-emergence	6,11,14	<u>Rhizobium leguminosarum</u>	Acifluorfen 0.1,0.4,1.6
7 19.11.79-13.3.80	White Clover	Kent Wild	10 - 5	2-3 Leaves	4,10	-	Fluazifop-butyl 0.1,0.4,1.0

ResultsALLOXYDIM SODIUMCode Number NP48, BAS 9021Trade names Fervin, Clout.Chemical name 2-(1-allyloxyaminobutylidene)-5,5-dimethyl-4-methoxycarbonyl cyclohexane-1,3-dione (sodium salt)Source Nippon Soda Co. Ltd., Agrochemicals Department, 221 Ohtemachi, Chiyoda-ku, Tokyo, Japan.Formulation 75% w/w a.i. water soluble powder.Spray volume 345 l ha⁻¹.Spray rates used in experiments 2 kg a.i. ha⁻¹.Information available and suggested use

The suggested post-emergence use is for control of grass weeds. Broad-leaved weeds are resistant. Dosage for annual weeds, 1.0 - 2.0 kg a.i. ha⁻¹; for perennials 1.5 - 2.5 kg a.i. ha⁻¹, two application times being advisable. Recommended for use in sugar beet, cotton and rape. Other tolerant crops include:- groundnut, potato, beans, sunflower, tobacco, vines and various vegetables. Also active pre-emergence but not recommended due to its short persistence in soil. See also Richardson and Parker (1978).

ResultsExperiment 1

Table 3. Effect of growth stage at spraying on response of peas to alloxymid sodium (2kg a.i. ha⁻¹).

Sample time (wks after spraying)	Spray Stages					
	3 leaf			6 leaf		
	3	6	10	3	6	10
Shoot dry wt.∇	94	88	76*	96	ND	87
Root dry wt.∇	93	89	78*	89	ND	96
Nodule index	ND	10	8.5	10	ND	13

∇ Results expressed as % of control

* Significant difference from control at $P < 0.05$

ND - Not Determined

Comments on ResultsExperiment 1 (Table 3)

Spraying at the 3 leaf stage decreased the root and shoot dry weights at both 3 and 6 week sample dates. Ten weeks after spraying the decreases were statistically significant. The reduction in root dry weight at 10 weeks was accompanied by a small but noticeable reduction in nodulation. No significant weight decreases occurred with peas sprayed at the 6 leaf stage, and nodulation appeared to be slightly increased by the herbicide. Slight, non-significant reduction in pod yield was associated with the herbicide treatment at both growth stages.

Similar results were found in earlier experiments by Greaves *et al.*, (1978).

Experiment 2

Table 4. Effect of fertilizer level on the response of peas to alloxym sodium (2 kg a.i. ha⁻¹)

Fertilizer level (g kg ⁻¹ soil)		Sample time (weeks after spraying)		
		3	6	10
Shoot dry wt. ∇				
0		97	95	82*
0.2		95	93	91
0.8		89	84*	80
3.5		90	77*	95
Root dry wt. ∇				
0		90	73*	82*
0.2		84	66*	76*
0.8		88	90	72*
3.5		64*	61*	111
Pod dry wt. ∇				
0		ND	ND	184*
0.2		ND	ND	156
0.8		ND	ND	103
3.5		ND	ND	184
Nodule index				
0	Control	10	10	10
	Herbicide	9	8	9.5
0.2	Control	3	7	7.5
	Herbicide	2	5	6.5
0.8	Control	1	3	3.5
	Herbicide	1	4	2.5
3.5	Control	0	1	0.5
	Herbicide	0	0	0

∇ Results expressed as % of control

* Significant difference from control at P < 0.05

ND Not Determined

Comments on Results

Experiment 2. (Table 4)

Alloxym sodium treatment reduced shoot dry weight at all fertilizer levels, confirming the results following spraying at the 3 leaf stage in experiment 1. This effect increased with time after spraying up to the final assessment at 10 weeks, except at the 3.5 g kg⁻¹ fertilizer level when the maximum effect occurred at 6 weeks. As shoot dry weights varied appreciably, the only statistically significant effects occurred at 10 weeks (0 fertilizer) and 6 weeks (3.5 g kg⁻¹ fertilizer). Root dry weights were reduced by the herbicide at all fertilizer levels. The results were less variable than those of the shoots and, thus the effects were more frequently statistically significant.

Increasing the amount of fertilizer decreased nodulation markedly at all sample dates, and caused the replacement of large, pink, healthy nodules with small, white, ineffective nodules. The herbicide had only a slight additional effect on nodule density. Herbicide treatment appeared to increase pod yield at all fertilizer levels, but the effect was significant only when no fertilizer was used, it was also noted that at this fertilizer level the plants flowered earlier than all other treatments.

The plants grown at the 3.5 g kg^{-1} fertilizer level were very much larger than those grown at all other fertilizer levels. Consequently retention of the herbicide was affected, being 7 to 15 times higher than plants grown at lower fertilizer levels. Retained herbicide was removed from plants by washing with 0.1M NaOH. The alloxym sodium was partitioned into hexane and determined by gas liquid chromatography (pers. comm. E. G. Cotterill). Considering the very large increase in retention it is reassuring that the effects on plant growth and nodulation are not increased proportionally.

Conclusions

Alloxym sodium has only slight inhibitory effects on pea growth and nodulation. These effects are minimized or eliminated if the plants are sprayed at the 6 leaf stage, rather than the 3 leaf stage.

SETHOXYDIMCode number NP 55, ARD 34/02Chemical name 2-(N-ethoxybutyrimidoyl)-5-(2-ethylthiopropyl)-3-hydroxy-2-cyclohexen-1-one.Source May & Baker Ltd., Ongar Research Station, Fyfield Road, Ongar, Essex, UK.Formulation 19.3% w/v a.i. emulsifiable concentrateSpray volume 370 l ha⁻¹.Spray rates used in experiments 0.1, 0.4, 1.0, and 1.6 kg a.i. ha⁻¹.Information available and suggested uses.

Used to control grass weeds in broad leaved crops. The herbicide is absorbed by both roots and shoots, and because of its short persistence its main use is post-emergence. Efficiency and speed of action is increased by warm wet conditions, and decreased by cold dry conditions. Tolerant crops include peas, field beans, dwarf french beans, onions and several other vegetables.

RESULTSExperiment 3.Table 5. Effect of dose rate of sethoxydim on growth and nodulation of peas

Dose (kg ha ⁻¹)	Sample time (weeks after spraying)		
	3	6	8
	<u>Shoot dry wt. ∇</u>		
0.4	102	94	94
1.0	91	95	101
1.6	99	74	88
	<u>Root dry wt. ∇</u>		
0.4	98	103	97
1.0	92	117	110
1.6	93	101	92
	<u>Nodule index</u>		
0.4	10	10	5
1.0	6	8	6
1.6	3	4	4

∇ Results are expressed as % of control

Comments on ResultsExperiment 3 (Table 5).

There was no significant decrease in root and shoot dry weights at any of the sample times. Nodulation was markedly reduced at the higher herbicide concentrations, and by the final assessment similar reductions were found in the lower doses. There was no effect on the pod yield.

Experiment 4.Table 6. Effect of growth stage at spraying on response of peas to sethoxydim (1 kg ha⁻¹)

Sample time (weeks after spraying)	Growth stages at spraying			
	3 leaf		6 leaf	
	3	5	3	5
Shoot dry wt. ∇	126	106	94	87
Root dry wt. ∇	110	104	97	101
Nodule Index	8	4	8	7

∇ Results are expressed as % of control

Comments on ResultsExperiment 4. (Table 6).

The plants in this experiment were grown in soil with no added fertilizer, and the experiment had to be terminated early due to severe growth of downy mildew on all the peas. Thus, the results must be interpreted with caution. However, it does appear that there was no harmful effect of the herbicide on the plant growth, although after 5 weeks nodulation appeared to be inhibited in plants sprayed at the 3 leaf stage.

Experiment 5Table 7 The effect of sethoxydim at 3 dose rates on peas

Herbicide dose (kg ha ⁻¹)	Sample time (weeks after spraying)	
	3	7
	<u>Shoot dry wt. ∇</u>	
0.4	101	107
1.0	97	115
1.6	99	103
	<u>Root dry wt. ∇</u>	
0.4	96	112
1.0	93	111
1.6	93	105
	<u>Pod dry wt. ∇</u>	
0.4	ND	114
1.0	ND	116
1.6	ND	115
	<u>Nodule index</u>	
0.4	9	6
1.0	7	6
1.6	6	8

∇ Results expressed as % of control
ND Not determined

Table 8 The effect of sethoxydim at 3 dose rates on dwarf french beans

Herbicide dose (kg ha ⁻¹)	Sample time (weeks after spraying)	
	3	6
	Shoot dry wt. ∇	
0.4	106	101
1.0	94	99
1.6	103	95
	Root dry wt. ∇	
0.4	125	105
1.0	118	106
1.6	118	87

∇ Results expressed as % of control

Table 9. The effect of sethoxydim at 3 dose rates on field beans

Herbicide dose (kg ha ⁻¹)	Sample time (weeks after spraying)		
	+ VITAX		- VITAX
	4	8	8
	Shoot dry wt. ∇		
0.4	88	84*	93
1.0	82	91	94
1.6	72*	79*	96
	Root dry wt. ∇		
0.4	89	83	81
1.0	72*	83	81
1.6	77*	64*	74
	Nodule index		
0.4	3	3	7
1.0	1	5	5
1.6	5	1	3

∇ Results expressed as % of control

* Significant difference to control at $P < 0.05$

Comments on Results

Experiment 5 (Tables 7-9)

Peas (Table 7)

The peas sprayed with a high concentration of sethoxydim showed signs of spray damage, scorching of the lower leaves. By the final harvest all the peas were badly infected with mildew, so care must be taken in interpretation of the results. Sethoxydim had little if any effect on the root and shoot dry weights

of the peas at either harvest time. Nodulation was reduced, but pod yield increased slightly with herbicide dose.

Dwarf french beans (Table 8)

Growth was poor, and consequently the plants were harvested early. There was little effect of sethoxydim on the shoot dry weights, and only a slight stimulation of root growth at the 3 week sampling. The plants did not nodulate, despite being inoculated with Rhizobium phaseoli, and being grown in soil not amended with fertilizer.

Field beans (Table 9)

The main part of the experiment consisted of field beans grown in soil amended with Vitax QS3, and a smaller section in soil without Vitax. Initial observation of all the beans showed that the plants sprayed with the highest dose of sethoxydim showed signs of spray damage, lower leaves being scorched and dying off.

The growth of plants in soil with fertilizer were inhibited by sethoxydim with reductions in both the root and shoot dry weights. Nodulation was severely inhibited.

The beans grown in soil with no Vitax showed little effect of sethoxydim on the shoots, but there were reductions in the root dry weights, and a reduction in nodule index, increasing with dose.

There were more nodules on beans grown in soil with no fertilizer than those grown with fertilizer.

Herbicide Selectivity Evaluation (Richardson, West and Parker, 1980)

Table 10. Effects of sethoxydim on fenugreek, field bean, pea and white clover.

Herbicide dose (kg ha ⁻¹)	Fenugreek	Field bean	Pea	Clover
		<u>Shoot dry wt. ∇</u>		
0.1	56	ND	96	114
0.4	75	65	79	99
1.6	73	86	73	98
		<u>Root dry wt. ∇</u>		
0.1	59	ND	87	101
0.4	74	67	76	86
1.6	75	106	73	79
		<u>Nodules</u>		
0.1	5	ND	7	10
0.4	11	6	5	10
1.6	6	5	8	10

∇ Results expressed as % control Sampled 5 weeks after spraying.

Herbicide Selectivity Evaluation (Richardson, West and Parker, 1980)Table 11. Effect of different types of application of sethoxydim on dwarf french beans.

Herbicide application:-	A	B	C
Herbicide dose (kg ha ⁻¹)	Shoot dry wt. ∇		
0.4	122	91	106
1.6	130	98	71
	Root dry wt. ∇		
0.4	54	136	123
1.6	82	133	100

∇ results expressed as % control. Sampled 8 weeks after spraying.

A - pre-emergence; soil surface sprayed.

B - pre-emergence; herbicide incorporated into soil.

C - post emergence.

Comments on ResultsHerbicide Selectivity Evaluation (Tables 10 and 11) (Richardson, West & Parker, 1980)

Work done in routine herbicide tests included experiments on the effect of sethoxydim on several legumes. Root and shoot dry weights of peas were reduced by up to 27% at the highest herbicide concentration and nodulation was also reduced. Fenugreek and field beans were affected similarly except at the highest concentration where the root dry weights of field beans were slightly increased. There was little effect on clover.

Different application methods of sethoxydim were tested on dwarf french beans. Spraying directly onto the soil surface prior to emergence resulted in a marked reduction of root dry weight, particularly at the lowest dose, but there was no increase in shoot weight. In contrast, when the herbicide was incorporated into the soil after pre-emergence application, root dry weight was increased and shoot weights were scarcely affected. Post-emergence treatment also resulted in an increase of root dry weight, but only at the low dose. There was some reduction of the shoot growth at the high dose. Again, this species did not nodulate.

Conclusions

Effects of sethoxydim on plant growth were generally small and variable. Nodulation was decreased, particularly by the higher herbicide dose rates, in all the legume species tested except clover.

ACIFLUORFEN SodiumCode number RH 6201Trade name BlazerChemical name Sodium 5-[2-chloro-4-(trifluoromethyl)phenoxy]-2-nitrobenzoateSource Rohm & Haas (UK) Ltd., Lennig House, 2, Masons Avenue, Croydon, Surrey, CR9 3NB UK.Formulation 48% w/v a.i. aqueous concentrate, sodium salt.Spray volume 437 l ha⁻¹.Spray rates used in experiments 0.1, 0.4 and 1.6 kg ha⁻¹.Information available and suggested uses

A contact and residual herbicide, pre- and post-emergence control of most broadleaf weeds in all large seeded legumes (soyabean, peas, beans, peanuts). Current problem weeds such as cocklebur, velvetleaf, morningglory are susceptible at 0.56 kg ha⁻¹. Its chemical, physical and biological properties have been reported by Johnson *et al* 1978.

ResultsExperiment 6Table 12. The effect of a pre-emergence spray of acifluorfen on growth of peas

Herbicide dose (kg ha ⁻¹)	Sample date (wks after spraying/planting)		
	6	11	14
	<u>Shoot dry wt.∇</u>		
0.1	110	88	98
0.4	129	100	98
1.6	116	104	118
	<u>Root dry wt.∇</u>		
0.1	107	111	64
0.4	141*	103	82
1.6	122	60	90
	<u>Pod dry wt.∇</u>		
0.1	ND	ND	97
0.4	ND	ND	92
1.6	ND	ND	93

∇ Results expressed as % of control

* Significant difference from control at P < 0.05

ND Not Determined

Comments on ResultsExperiment 6. (Table 12)

At the early sample date there was a tendency towards stimulation of both root and shoot dry weights, the stimulation of root dry weight being significant at 0.4 kg ha⁻¹. At the final harvest, this effect had generally reversed into a slight reduction. Nodulation was poor throughout the experiment, despite inoculation with the appropriate rhizobium culture, thus nodulation was not assessed. Pod dry weights were only slightly reduced.

Herbicide Selectivity Evaluation (Richardson, West and Parker, 1979)Table 13. Effects of acifluorfen on growth of dwarf french beans and field beans.

Herbicide dose (kg ha ⁻¹)	DWARF FRENCH BEANS	FIELD BEANS
	<u>Shoot dry wt. ∇</u>	
0.1	87	89
0.4	83	87
1.6	52	66
	<u>Root dry wt. ∇</u>	
0.1	105	116
0.4	79	86
1.6	41	68

∇ Results expressed as % of control
Plants assessed 6 weeks after spraying.

Comments on resultsHerbicide Selectivity Evaluation (Table 13)

This experiment showed a reduction in root and shoot dry weights in both dwarf french beans and field beans, the effect increased with dose in both cases. No nodulation assessment was made due to poor nodulation of both species.

Peas were also tested in this experiment but only at 1.6 kg ha⁻¹, and at this dose there was a reduction in root and shoot dry weight to 84 and 98% of control respectively. Nodulation was also reduced to an index value of 6.

Conclusions

In general acifluorfen had little significant effect on pea growth, although nodulation was reduced. However, the reduction in growth of dwarf french beans and field beans may be of some concern, and will need further investigation.

FLUAZIFOP-BUTYL

Code number PP009
 Trade name Fusilade
 Chemical name Butyl 2-[4-(5-trifluoromethyl-2-pyridyloxy)phenoxy] propionate
 Source ICI Plant Protection Ltd., Jealotts Hill Research Station, Bracknell,
 Berks RG12 6EY UK.
 Formulation 25% w/v a.i. emulsifiable concentrate.
 Spray volume 370 l ha⁻¹.
 Spray rates used in experiments 0.1, 0.4, 1.0 and 1.6 kg ha⁻¹.
Information available and suggested uses

Selective post-emergence herbicide for control of annual and perennial grass weeds in broad-leaved crops (sugar beet, winter oil seed rape, field beans, cabbage, potatoes). Weeds controlled include; rye grass, wild oats, black grass, barren brome and couch. Treatment is only necessary on the appearance of the weeds.

ResultsExperiment 7

Table 14. The effect of fluazifop-butyl on white clover

Herbicide dose (kg ha ⁻¹)	Sample time (weeks after spraying)	
	+ VITAX	- VITAX
	4	10
	<u>Shoot dry wt.∇</u>	
0.1	98	93
0.4	89	ND
1.0	72*	87
1.6	86	89
	<u>Root dry wt.∇</u>	
0.1	94	95
0.4	86	ND
1.0	70*	97
1.6	85	103
	<u>Nodule index</u>	
0.1	1	5
0.4	3	ND
1.0	1	5
1.6	3	5

∇ Results expressed as % control

* Significant difference from control at $P < 0.05$

ND Not determined

Comments on ResultsExperiment 7 (Table 14).

In soil amended with fertilizer, fluazifop-butyl caused small reductions in both root and shoot dry weights at all concentrations. This effect was

statistically significant at the 1.0 kg ha^{-1} dose 4 weeks after spraying, but by 10 weeks the plants had recovered. In the absence of fertilizer the effects at 10 weeks were similar.

In fertilized soil, nodulation, was initially severely affected by all doses but showed some recovery, after 10 weeks. In contrast, in unfertilized soil nodulation was markedly reduced by all doses 10 weeks after treatment.

Herbicide Selectivity Evaluation (Richardson, West and Parker, 1980)

Table 15. Effect of post emergence treatment with fluazifop-butyl on growth and nodulation of white clover and dwarf french beans.

Herbicide dose (kg ha^{-1})	CLOVER	DWARF FRENCH BEANS
0.1	107	106
0.4	147	119
1.6	109	66
		<u>Root dry wt. ∇</u>
0.1	129	94
0.4	160	105
1.6	148	90
		<u>Nodule index</u>
0.1	9	ND
0.4	6	ND
1.6	4	ND

∇ Results expressed as % control
Plants assessed 5 weeks after spraying

Table 16. Effect of post emergence treatment with fluazifop-butyl on growth and nodulation of fenugreek, pea and field bean.

Herbicide dose (kg ha ⁻¹)	<u>FENUGREEK</u>	<u>PEA</u>	<u>FIELD BEAN</u>
		<u>Shoot dry wt. ∇</u>	
0.1	103	81	86
0.4	132	89	89
1.6	116	112	88
	<u>Root dry wt. ∇</u>		
0.1	91	87	96
0.4	113	74	90
1.6	98	89	83
	<u>Nodule Index</u>		
0.1	3	7	5
0.4	1	8	7
1.6	3	10	9

∇ Results expressed as % of control
Plants assessed 5 weeks after spraying

Table 17. Effect of fluazifop-butyl on the growth of dwarf french beans

Herbicide Treatment:-	A	B	C	D
	<u>Shoot dry wt. ∇</u>			
Herbicide dose (kg ha ⁻¹)				
0.1	97	118	80	107
0.4	74	100	99	87
1.6	88	82	79	87
	<u>Root dry wt. ∇</u>			
0.1	101	119	67	82
0.4	75	112	95	72
1.6	104	94	73	70

∇ - Results expressed as % of control
A - Post-emergence, foliar spray only
B - Post-emergence, soil drench
C - Pre-emergence, soil surface sprayed
D - Pre-emergence, herbicide incorporated into soil
Plants assessed 8 weeks after treatment

Comments on Results

Herbicide Selectivity Evaluation (Table 15-17)

Earlier work at WRO (Richardson, West and Parker, 1980) (Table 15) showed that although nodulation of clover was progressively reduced by increasing dose rate both shoot and root growth showed increases due to herbicide treatment.

Dwarf french beans showed only very slight and variable growth responses, except for a 34% decrease in shoot dry weight caused by 1.6 kg ha^{-1} fluazifop-butyl. Nodulation did not occur with this species.

Fluazifop butyl increased shoot dry weights of fenugreek and caused small and variable effects on root growth. With peas and field bean, both root and shoot dry weights were generally reduced.

Nodulation was severely inhibited in fenugreek at all doses, despite only slight effects on roots and shoots. In contrast nodulation of pea and field bean was only slightly reduced, the effect being greatest at the lowest dose.

Assessment of the effect of application method (Table 17) showed that, in general, root and shoot growth were reduced by pre-emergence treatments, particularly when the herbicide was not subsequently incorporated. Only at 0.4 kg ha^{-1} did post-emergent treatment cause any notable reduction.

Conclusions

Fluazifop-butyl effects on growth of the species tested were generally small. However, the increased shoot growth of fenugreek and clover is of interest. Nodulation of fenugreek and clover can be markedly reduced, though, in the case of clover at least, this may recover some time after spraying (Table 14).

Although pre-emergence treatment of dwarf french bean caused damaging effects, this is at present of little practical importance as the herbicide is only recommended for post-emergence use.

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ABBREVIATIONS

ångström	Å	freezing point	f.p.
Abstract	Abs.	from summary	F.s.
acid equivalent*	a.e.	gallon	gal
acre	ac	gallons per hour	gal/h
active ingredient*	a.i.	gallons per acre	gal/ac
approximately equal to*	≈	gas liquid chromatography	GLC
aqueous concentrate	a.c.	gramme	g
bibliography	bibl.	hectare	ha
boiling point	b.p.	hectokilogram	hkg
bushel	bu	high volume	HV
centigrade	C	horse power	hp
centimetre*	cm	hour	h
concentrated	concd	hundredweight*	cwt
concentration	concn	hydrogen ion concentration*	pH
concentration x time product	ct	inch	in.
concentration required to kill 50% test animals	LC50	infra red	i.r.
cubic centimetre*	cm ³	kilogramme	kg
cubic foot*	ft ³	kilo (x10 ³)	k
cubic inch*	in ³	less than	<
cubic metre*	m ³	litre	l.
cubic yard*	yd ³	low volume	LV
cultivar(s)	cv.	maximum	max.
curie*	Ci	median lethal dose	LD50
degree Celsius*	°C	medium volume	MV
degree centigrade	°C	melting point	m.p.
degree Fahrenheit*	°F	metre	m
diameter	diam.	micro (x10 ⁻⁶)	μ
diameter at breast height	d.b.h.	microgramme*	μg
divided by*	÷ or /	micromicro (pico: x10 ⁻¹²)*	μμ
dry matter	d.m.	micrometre (micron)*	μm (or μ)
emulsifiable concentrate	e.c.	micron (micrometre)*†	μm (or μ)
equal to*	=	miles per hour*	mile/h
fluid	fl.	milli (x10 ⁻³)	m
foot	ft	milliequivalent*	m.equiv.
		milligramme	mg
		millilitre	ml

† The name micrometre is preferred to micron and μm is preferred to μ.

millimetre*	mm	pre-emergence	pre-em.
millimicro* (nano: $\times 10^{-9}$)	n or mp	quart	quart
minimum	min.	relative humidity	r.h.
minus	-	revolution per minute*	rev/min
minute	min	second	s
molar concentration*	M (small cap)	soluble concentrate	s.c.
molecule, molecular	mol.	soluble powder	s.p.
more than	>	solution	soln
multiplied by*	x	species (singular)	sp.
normal concentration*	N (small cap)	species (plural)	spp.
not dated	n.d.	specific gravity	sp. gr.
oil miscible concentrate	o.m.c. (tables only)	square foot*	ft ²
organic matter	o.m.	square inch	in ²
ounce	oz	square metre*	m ²
ounces per gallon	oz/gal	square root of*	$\sqrt{\quad}$
page	p.	sub-species*	ssp.
pages	pp.	summary	s.
parts per million	ppm	temperature	temp.
parts per million by volume	ppmv	ton	ton
parts per million by weight	ppmw	tonne	t
percent(age)	%	ultra-low volume	ULV
pico (micromicro: $\times 10^{-12}$)	p or pp	ultra violet	u.v.
pint	pint	vapour density	v.d.
pints per acre	pints/ac	vapour pressure	v.p.
plus or minus*	+ -	<u>varietas</u>	var.
post-emergence	post-em	volt	v
pound	lb	volume	vol.
pound per acre*	lb/ac	volume per volume	v/v
pounds per minute	lb/min	water soluble powder	w.s.p. (tables only)
pound per square inch*	lb/in ²	watt	w
powder for dry application	p. (tables only)	weight	wt
power take off	p.t.o.	weight per volume*	w/v
precipitate (noun)	ppt.	weight per weight*	w/w
		wettable powder	w.p.
		yard	yd
		yards per minute	yd/min

* Those marked * should normally be used in the text as well as in tables etc.



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